## POTENTIAL OF MACRO-INVERTEBRATES IN BIOACCUMULATION OF HEAVY METALS – A REVIEW

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**Abstract** – Benthic macro-invertebrates are the most appropriate and ideal organisms for bio-indicators of biodegradable organic pollution in surface water bodies. Apart from this specific feature many researchers have found that some of the orders (Molluscs, Annelides, Coleoptera, Polychaeta and Oligochaeta) of benthic macroinvertebrates can also accumulate heavy metals. This review highlights the case studies of different groups of macro-invertebrates for heavy metal bioaccumulation, which help to understand the fact that macroinvertebrates can serve as potential bioindicators of heavy metal pollution.

### INTRODUCTION

Heavy metal pollution is a major global concern due to its non-degradable nature in the environment. It creates soil as well as water pollution that accumulates into aquatic organisms. This accumulation further travels into higher organisms through the food chain and causes harmful health effects to humans and animals (Cheng, 2003). Benthic macro-invertebrates are very useful bioindicators of organic pollution. According to their sensitivity towards organic pollution, the taxonomic orders have been categorized into pollution sensitive, moderately tolerant, and highly tolerant groups. Pollution sensitive group includes Ephemeroptera, Trichoptera and Plecoptera orders that can survive only in the waterbody where dissolved oxygen is high and the less biodegradable organic pollutants (Ghosh and Biswas, 2017)

Contrary to this, a highly tolerant group includes few families of Oligochaeta and Diptera orders. They make several adaptations in their body, due to which they can survive even in significantly less/nil dissolved oxygen conditions and very high amounts of organic pollutants. Adaptations such as siphon in the Syrphidae family, the red pigment in Chironomidae worms, and cuticle layers in oligochaetes make them tolerant in highly polluted environments (Porst*et al.*, 2012). The distribution of benthic macroinvertebrates was widely studied as biomonitors and the rivers were bio-mapped in European countries based on the presence/absence of these macro-invertebrates.

Many have developed bio-alarm systems based on the characteristics of benthic macroinvertebrates to monitor sudden changes in water bodies due to pollution. The heavy metals are available in sediment, or water becomes available for the benthic macroinvertebrates. These invertebrates accumulate them into their different body parts such as tissues, organs, etc. Bettles belonging to Carabidae family have been found to segregate heavy metals and convert them into less active forms through different detoxification mechanisms (Hopkin, 1989). They might have a system to accumulate metals in the hindgut wall (Butovsky, 2011). The concentrations of heavy metals accumulation depend on the contamination in the environment and vary with the organisms. Heavy metals such as Zn, Cu might be required for their metabolism that was significantly less amount, but they are bound to uptake other heavy metals into their bodies, such as cadmium, arsenic, lead, etc., due to their high concentrations in the surrounding environment (Table 1). However, it has been studied that in living organisms, due to the internally metabolically available heavy metals, all vital or non-vital metals become lethal to the organism despite their concentration (Rainbow, 2007). These macroinvertebrates can serve as bioindicators to

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enumerate the bioavailable amount of pollutants in water bodies. (Zulkifli et al., 2012). Research studies have shown that molluscs can store high concentrations of heavy metals and survive in adverse environments (Amiard et al., 1986; Usero et al., 1997; Joiris and Azokwu, 1999; Ananthan et al., 2006; Kesavan et al., 2013). The reason may be that molluscs feed on phytoplankton and bottom sediments, containing high levels of pollutants (Luoma and Jenne, 1976). The prime reason of heavy metal exposure may be the diet (Poteat and Buchwalter, 2014) instead of direct aqueous exposure (Xie et al., 2010). A correlation pattern has been observed between levels of Rare Earth Elements (REE) and the density of macroinvertebrates category belonging to collectorgatherers which defines that predatory organisms are more efficient in accumulating REEs than nonpredatory invertebrates and therefore, collectorsgatherers can be useful organisms to trace elements in freshwater ecosystems.

However, there is a vast gap in the literature; some successful attempts have been made to understand the bioaccumulation of heavy metals in macro-invertebrates (Table 2). Thus, the present review highlights such research studies, which will also help explore the use of macroinvertebrates in heavy metal bioaccumulation.

# Methodology for analysis of bioaccumulation studies in macro-invertebrates

The common methodology was adapted in all the available literatures with modifications according to the type of organisms. The water/soil/sediments and macro-invertebrates samples from the contaminated sites were collected. The macrobenthos samples are washed thoroughly in double distilled water to remove the debris or soil particles, further, fixed in 4% formaldehyde to prevent decomposition of organisms. Biota is then identified to study the different taxonomic groups present at the site and after laboratory sorting theyare kept in 70% alcohol (ISO11047/1998). For heavy metal analysis, macrobenthos samples (shells of molluscs and soft tissues in case of other macrobenthos) are first dried in oven at a temperature of 60%C. Dried/powdered tissues (0.1 g) were after wards acid digested with 2 mL of a mixture of nitric and perchloric acid (ratio 9:1) (Julshmamn et al., 1982). Digested samples are filtered through Whatman filter paper no.41. The metal concentrations were determined using AAS (Dar et al., 2018).

The heavy metal concentration in surrounding water/sediment and organisms' body are compared to understand the Bioaccumulation Factor (BAF) according to Klavins *et al.* (1998) (Equations 1 and 2):

 $BAF_{Wv} = M_{benthos} / M_{wat} (1)$  $BAFSv = M_{benthos} / M_{sed} (2)$ 

where  $M_{benthos}$  is the metal amount in the benthos;  $M_{wat}$  is the metal amount in the water, and  $M_{sed}$  is the amount in the sediment.

#### **Case studies**

Latif et al. (2011), explored the potential of oligochaete Eiseniellatetraedra as a bioindicator of cadmium (Cd) and lead (Pb) contamination in Iran. The concentration of heavy metals was analysed in soil and earthworm tissues from two locations; Polur Falls and the riverside area of Abali. Pb and Cd concentrations in Earthworms collected from Abali were found to be higher than the earthworms collected from Polur (Pb: Polur=38.02±4.44 ppm, Abali=199.92±21.58 ppm; Cd: Polur=7.35±0.97 ppm, Abali=16.77±2.92 ppm). The accumulation of Pb and Cd in earthworms from Abali was 5 and 2 times higher, respectively than from Polur. Also, it was observed that Pb and Cd contents in earthworms were higher than in the surrounding soils at both stations. Hence, they recommended the use of *E*. tetraedra as a bioindicator of heavy metals contaminated locations (Latif et al., 2011).

Dange and Manoj (2015) reported that the polychaete worms were used for bioaccumulation from Purna river estuary, South Gujarat. Heavy metals Copper, Nickel, Zinc and Cadmium concentrations were determined in sediment and several tissues of polychaetes. It was observed that the body wall and gut are the major body parts in Nereididae family of polychaete used for bioaccumulation of copper. In the case of Mud crab (Rhithropanopeusharrisii), heavy metals Copper, Nickel and Zinc were accumulated in different body parts such as chelipeds, walking legs, carapace etc. except Cadmium, Cu and Zn are essential elements for crabs as they are required for different biological functions but in limited concentrations. (Dange and Manoj, 2015).

Carabidae beetles make the major part of soil bioindicators. It has been found that beetles have gut as the exceptionally important body part which allows uptake, transmission, accumulation and excretion of metals (Agnieszka *et al.*, 2016) while midgut restricts the toxic materials inside the body while eating (Rawi, 2011). Nasr and Khater (2020) examined their use as an indicator of heavy metal pollution in soil. The heavy metal (As, Hg, Al, Cd, Se, and Pb) concentrations were determined in soil and midgut of C. Chlorostictum beetle. Also, physicochemical properties of the sampling site were also studied. The data reflected that As was found to be the most accumulated metal (11.858 µg/ g<sup>-1</sup>) in the midgut of insects collected from polluted site in comparison to reference site this may be due to the higher concentration of arsenic in the zone. Concentrations of other metals found in the insect midgut were in the order as Al>Pb>Cd>Hg>Se. Transmission Electron Microscopic (TEM) results showed that heavy metals caused ultrastructural changes in the midgut epithelial layer of the ground beetles, collected from control and contaminated sites. Apart from structural changes correlation studies of heavy metals concentrations among soil and insects midgut were also done among the which reflected highest correlation for As and lowest observed was for Al (Nasr et al., 2020)

Hydrophilidae is one of the widely distributed families of order Coleoptera, it haswell developed taxonomy. Since, Hydrophilidae larvae needvarious environmental surroundings, they can be suitable as heavy metal bioindicators (Fikáèek *et al.*, 2010).

The potential of Hydrophilidae was explored by Aydoðanet. al. (2017) they assessed the heavy metal concentrations in sediment, water, fish (Chondrostomaregium) tissues, and some aquatic insects (Coleoptera: Hydrophilidae) concentrations in two study areas in Erzurum, Turkey (Aºkale and Dumlu sites). In fish tissues, 11 metals (Al, Cr, Mn, Fe, Ni, Cu, Zn, As, Cd, Hg, Pb) were analysed, and in insects (Laccobius (Dimorpholaccobius) syriacus, Laccobius (D.) imulatrix, Laccobius (D.) bipunctatus, Enochrus (Lumetus) bicolor, Enochrus (L.) fuscipennis, Enochrus (L.) halophilus, and Helochareslividoides of hydrophilidae family, 14 elements (Ti, V, Cr, Mn, Fe, Co, Ni, Cu, Zn, As, Se, Sr, Br, Pb) were measured. They concluded that insects were affected by heavy metal contamination in the surrounding area and could accumulate higher concentrations of metals in comparison to fishes (Aydoðan et al., 2017).

Molluscshave a capacity to accumulate heavy metals differentially in the soft tissues and calcium carbonate shells of the bivalvesand thus, can be very good bio-indicators of heavy metal pollution (Szefer and Szefer, 1990). However, most studies suggest the majority of accumulation occurs in the soft tissues but based on some studies, shells offer more accurate information about heavy metals accumulation. (Carriker *et al.*, 1982; Thorn *et al.*, 1995; Huanxin *et al.*, 2000; Yasoshima and Takano, 2001). Also, due to their long-life span (for example-*Margaritiferamargaritifera* and *Velesunioambiguus*, is upto100 & 30 years respectively), they can provide long-term impact of pollutants on aquatic bodies. (Carell *et al.*, 1987; Walker, 1992)

Dar et al. (2018) studied various trends of heavy metals mineralisation in some species of bivalves and molluscs which collected from the Timsah and Great Bitter. The capability of molluscs to biomineralize heavy metals into their shells was studied and depicted as Bio-mineralisation factor (BMF). At Timsah lake, Cardiumpapyraceum and Paratapesundulatus each showed highest BMFM ratios (0.95) of Ni. For Pb, highest BMFM ratio was found to be 0.92 in Cardiumpapyraceum. At the Great Bitter Lake, high Fe biomineralization was observed in Modiolusauriculatus and the Bulla ampulla within their shells in comparison to the other studied. Brachidontespharaonis accumulated highest Cd and Cu and similarly, Fusinus sp. accumulated highest concentrations of Ni and Zn. At the Timsah Lake, the accumulation of heavy metals was found in the following order Fe >Mn>Pb> Ni > Zn > Cu > Cd and at the Great Bitter Lake it was in following order Fe >Mn>Pb> Ni > Cd > Zn > Cu (Dar *et al.*, 2018).

Four bivalve molluscs (*Sanguinolariaacuminata, Anadaragranosa, Meretrixmeretrix, and Pelecyoratrigona*) were collected from collected from intertidal regions of the Sunderban mangrove wetland, north-eastern part of the Bay of Bengal and accumulation of heavy metals (Cu, Pb, Cd, Zn, and Hg) was studied by Sarkar *et al.* (2008). Contrary to many other research studies, shells have less accumulation capability than other soft body tissues

**Table 1.** Major Organs for metal accumulation in Macroinvertebrates

S.No.	Macroinvertebrates	Major organs for metal accumulation	
1.	Molluscs	Shell and soft body tissues (gill, visceral mass, mantle, podium, shell, adductor muscle )	
2.	Insects (beetles)	Gut, mid gut	
3.	Annelids	Gut	
4.	Crustacea	Chelipeds, Walking legs, carapace	

specifically, gills and mantle. The reason for high accumulation might be the ion exchange mechanism of the mucous layer layering these tissues. The higher accumulation of heavy metals into these bivalves may create harmful health effects for humans as they are consumed as seafood. On the basis of the results, it was observed that heavy metal accumulation varies with the species and affected by many factors such as size, age, sex and metal speciation, salinity etc. Also, each species of marine macroinvertebrates adopt different mechanism for each metal. Maximum accumulation of Cu and Zn, 175.14 and 2382 µg/g dry weight respectively in visceral mass of S. acuminata (bigger) at Gangasagar and in gills of A. granosa (larger at Jharkhali), which might be due to the fact that these two are essentially required in most of the metabolic processes of these bivalves (Greenfield et al., 2003; Paez-Osuna et al., 2002). S. acuminata could accumulate Pb 46.34 µg/g dry weight in the gill. An overall trend of organ specificity against accumulation of metals is observed and the sequence is as follows: gill> visceral mass> mantle> podium >shell >adductor muscle (Sarkar et al., 2008).

Bio-sorption of heavy metals (Cd, Pb, Zn) using bivalve molluscs was carried out by Mahendra *et al.* (2020). Shells were dried and crushed, then, shell powder was used to understand the sorption capacity of bivalve shells for Pb<sup>2+</sup>, Cd<sup>2+</sup>, and Zn<sup>2+</sup> was studied. Ion exchange mechanism Maximum concentration 489 mg/g for Pb 460 mg/g for Cd and 450 mg/g for Zn was obtained by seashell powder. The seashells are made up with CaCO<sub>3</sub> and present in calcite and aragonite forms (Vijayaraghavan et al., 2011, Afroze and Sen, 2018). Bio-sorption process was found to be influenced by temperature and pH. Sorption process was found to decrease with increase in temperature due to the damage of active binding sites. The effective pH range for bio-sorbent was found to be in range from 6-8. The metal ions replace the calcium ions present of shell surface through ion exchange mechanism, indicating the sorption process of metal uptake rather than adsorption (Mahendra et al., 2020). Shahzad et al. (2017), conducted a similar study on removing Pb (II) ions using shell biomass of cylindrical paper shell mussel (Anodontoidesferussacianus). Results indicated that the 200 mg/L of Pb (II) was removed through biosorption. The effect of some physicochemical parameters such as pH, contact time, temperature and biosorbent concentration etc. on biosorption process of A. ferussacianus shell was evaluated. Ideal pH and temperature were found to be 6 and 30°C respectively. The occurrence of some functional groups (-OH, NH, C=O and S=O) on the surface of bivalve mollusks helps in surface binding mechanisms of metals (Liu et al., 2009).

In addition to above mentioned studies, Bhalachandra *et al.*(2011) studied the heavy metal

Table 2. Common heavy metals and macroinvertebrates used in the bioaccumulation studies

Order	Name of Macro-invertebrates	Heavy metals studie	ed References
Oligochaeta	Eiseniellatetraedra	Zn, Cu	Latif et al. (2011)
Polychaeta	Nereididae	Cd, Pb,	Dange and Manoj (2015)
Mollusca	Brachidontespharaonis	Cd, Cu	Dar <i>et al.</i> (2018)
	Cardiumpapyraceum, Paratapes undulates	Ni, Pb	
	Modiolusauriculatus, Bulla ampulla	Fe	
	Fusinus sp.	Ni, Zn	
	Sanguinolariaacuminata, Anadaragranosa	Cu,Zn	Sarkar <i>et al</i> . (2008)
	Anodontoidesferussacianus	Pb	Liu et al. (2009)
	Lamellidenscorrianus	As, Pb	Bhalchandra et al. (2011)
	Lamellidensmarginalis	Cu, Hg	
	Parreysiacylindrica	Cd	
	Corbicula striatella	Zn	
	Indonaiacaeruleus	Cu, Hg	
Insecta	Calosomachlorostictum	As, Hg, Al, Cd,	Nasr and Khater (2020)
	Laccobius (Dimorpholaccobius) syriacus,	Se, and Pb	Aydoðan et al. (2017)
	(Laccobius (D.) imulatrix), Laccobius (D.)	Ti, V, Cr, Mn, Fe,	
	bipunctatus, Enochrus (Lumetus) bicolor,	Co, Ni, Cu, Zn, As,	
	Enochrus (L.) fuscipennis, Enochrus (L.) halophilus,	Se, Sr, Br, Pb	
	and Helochareslividoides		
Crustacea	Rhithropanopeusharrisii	Cu, Ni,Zn	Dange and Manoj, (2015)

uptake/accumulation into soft body tissues of the freshwater bivalves species namely, Corbicula Parreysiacylindrica, striatella, Parreysiacorrugata, Lamellidenscorrianus, Lamellidensmarginalis and Indonaiacaeruleus in laboratory conditions. The heavy metal concentrations used to acclimatize these bivalves were 0.1719 ppm for arsenic, 0.1411 ppm for cadmium, 0.033 ppm for copper, 1.50ppm for lead, 0.0443 ppm for mercury and 1.8589 ppm for zinc. The bivalves were exposed with heavy metals for up to 30 days in laboratory and accumulated concentration was analysed in bivalve tissues an dcontrol with a regular interval of 10, 20 and 30 days. Results indicated highest concentration of arsenic (37.9  $\mu$ g) and lead (1235.4  $\mu$ g) was accumulated by Lamellidenscorrianus, Lamellidensmarginalis accumulated highest concentration of copper (826.7 µg) and mercury (5.87 µg), Parreysiacylindrica accumulated the highest concentration of cadmium (182.62 µg) and the highest concentration of zinc (4139.2  $\mu$ g) was found in the per gram dry tissues of Corbicula *striatella*. Accordingly, bioconcentration factor (BCF) was also evaluated in same order as the heavy metal accumulation in bivalves. Based on the accumulation rate of different species, the *Lamellidenscorrianus* can be used as an indicator organism for the biomonitoring of arsenic and lead, Lamellidensmarginalis for copper and mercury, Parreysiacylindrica for cadmium and Corbicula striatella for zinc in freshwater ecosystem (Bhalchandra et al., 2011).

### CONCLUSION

The review highlights the potential of different aquatic as well as terrestrial species of macroinvertebrates in bioaccumulation of heavy metals. Since many research studies show different species of Mollusca group are most suitable as bioindicator species, it is suggested that their application in wastewater treatment technology as biosorbents may also be explored. Now, research should be focused on preparing a database of bioindicator species.

### REFERENCES

Afroze, S. and Sen, T.K. 2018. A review on heavy metal ions and dye adsorption from water by agricultural solid waste adsorbents. *Water Air & Soil Pollution*. 229 (7) : 1-50.

- Agnieszka, J., Bednarska, A., Laskowski, R., Elzÿbieta, P., Danuta, S., Zuzanna, S. and Olga, W. 2016. Metal toxicokinetics and metal driven damage to the gut of the ground beetle. Pterostichus Oblongopunctatus. *Environ. Sci. Pollut. Res.* 23 : 22047–22058.
- Amiard, J., Amiard-Triquet, C., Berthet, B. and Metayer, C. 1986. Contribution to the exotoxecological study of cadmium, lead, copper and zinc in the mussel Mytilusedulis. *Marine Biology*. 90 : 425–431.
- Ananthan, G., Sampathkumar, P., Palpandi, C. and Kannan, L. 2006. Distribution of heavy metals in velar estuary, southeast coast of India. *Journal of Ecotoxicology and Environmental Monitoring*. 16 (2) : 185–191.
- Aydoðan, Z.,ªiºman, T.,Ýncekara, Ü. and Gürol, A. 2017. Heavy metal accumulation in some aquatic insects (Coleoptera: Hydrophilidae) and tissues of *Chondrostomaregium* (Heckel, 1843) relevant to their concentration in water and sediments from Karasu River, Erzurum, Turkey. *Environ Sci Pollut Res.* DOI 10.1007/s11356-017-8629-x.
- Bhalachandra, W., Shinde, S. and Deshmukh, G. 2011. Bioaccumulation of Metal in Freshwater Pelecypod Molluscs under Experimental Condition. *Thebioscan*. 6 (4): 537-542.
- Butovsky, R.O. 2011. Heavy metals in carabids (Coleoptera, Carabidae). Zoo Keys. 100: 215–222.
- Carell, B., Forberg, S., Grundelius, E., Henrikson, L., Johnels, A., Lindh, U., Mutvel, H., Olsson, M., Svardstrom, K. and Westermark, T. 1987. Can mussel shells reveal environmental history? *Ambio*. 16 (1) : 2–10.
- Carriker, M.R., Swann, C.P. and Ewart, J.W. 1982. An exploratory-study with the proton micro-probe of the ontogenetic distribution of 16 elements in the shell of living oysters (*Crassostreavirginica*). *Mar. Biol.* 69 (3). 235–246.
- Cheng, S. 2003. Heavy metal pollution in China: origin, pattern and control. *Environ. Sci. Pollut. Res.*10 : 192-198.
- Dange, S. and Manoj, K. 2015. Bioaccumulation of Heavy Metals in Sediment, Polychaetes (Annelid) Worms, Mud Skipper and Mud Crab at Purna River Estuary, Navsari, Gujarat, India. *Int. J. Curr. Microbiol. App. Sci.* 4 (9) : 571-575.
- Dar, M. A., Belal, A. A. and Madkour, A. G. 2018. The differential abilities of some molluscs to accumulate heavy metals within their shells in the Timsah and the Great Bitter lakes, Suez Canal, Egypt. *Egyptian Journal of Aquatic Research*. 44 : 291–298.
- Fikáèek, M., Gentili, E. and Short, A.E.Z. 2010. Order Coleoptera, family Hydrophilidae. *Arthropod Fauna of the UAE*. 3 : 135–165.
- Ghosh, D. and Biswas, J.K. 2017. Efficiency of Pollution Tolerance Index (PTI) of macroinvertebrates in detecting aquatic pollution in an oxbow lake in India. *Univ. Sci.* 22 (3) : 237-261.

- Greenfield, B. K., Davis, J. A., Fairey, R., Roberts, C., Crane, D.B. and Ichikawa, Petreas, G.M. 2003. Contaminant Concentrations in Fish from San Francisco Bay, 2000, RMP Technical Report: SFEI Contribution 77. San Francisco Estuary Institute, Oakland, CA, USA.
- Hopkin, P.S. 1989. *Ecophysiology of Metals in Terrestrial Invertebrates*. Elseiver Applied Science Publishers Ltd. 366.
- Huanxin, W., Lejun, Z. and Presley, B.J. 2000. Bioaccumulation of heavy metals in oyster (*Crassostreavirginica*) tissue and shell. *Environ. Geol.* 39 (11) :1216–1226.
- ISO 11047/1998. Determination of Cadmium, Chromium, Cobalt, Copper, Lead, Manganese, Nickel and Zinc. Flame and Electrothermal Atomic Absorption Spectrometric Methods.
- Joiris, C.R. And Azokwu, M.I. 1999. Heavymetals in the bivalve Anadara (Senilia) senilis from Nigeria. *Marine Pollution Bulletin*. 38 (7): 618–622.
- Julshmamn, K., Ringdal,O., Slinning, K.E. and Braekan, O.R. 1982. Optimization of the Determination of Selenium in Marine Samples by Atomic Absorption Spectrometry: Comparison of a Flameless Graphite Furnace Atomic Absorption System with a Hydride Generation Atomic Absorption System. Spectrochimica Acta Part B: Atomic Spectroscopy. 37 : 6.473 – 482.
- Kesavan, K., Murugan, A., Venkatesan, & Vijay Kumar, B. 2013. Heavy metalaccumulation in molluscs and sediment from Uppanar Estuary, Southeast coast of India. An International Journal of Marine Sciences. 29(2). 15–21.
- Klavins, M., Briede, A., Parele, E., Rodinov, V. and Klavina, I. 1998. Metal accumulation in sediments and benthic invertebrates in Lakes of Latvia. *Chemosphere*. 36 (15) : 3043–3053.
- Latif, R., Malek, M. and MacKenzie, K. 2011.*Eiseniellatetraedra* (Savigny) (Oligochaeta: Lumbricidae) as a bioindicator of heavy metals. *Toxicological & Environmental Chemistry*. 93 (8): 1643– 1649.
- Liu, Y., Sun, C., Xu, J. and Li, Y. 2009. The use of raw and acid-pretreated bivalve mollusk shells to remove metals from aqueous solutions. *J. Haz. Mat.* 168:156-162.
- Luoma, S. And Jenne, E. 1976. Factors affecting the availability of sediment-bound cadmium to the estuarine, deposit-feeding clam, Macomabalthica. In: *Proceedings, Fourth National Symposium on Radioecology.* Ed. by Cushing, C. E. Jr. (ed.) Ecol. Soc. Am., Stroudsburg, Pa, 283–290.
- Mahendra, Ch., Sivakiran, R.R., Badrinarayana, K.A., Priya, L., Raj S. and Mamatha, M. 2020. Investigation of bivalve molluscan seashells for the removal of cadmium, lead and zinc Metal ions from wastewater streams. *Rasayan J. Chem*.13 (2) : 903-914.
- Nasr, E.E., Khater, Z. Z., Zelenakova, M., Vranayova, Z. and Abu-Hashim, M. 2020. Soil Physicochemical Properties, Metal Deposition, and Ultrastructural

Midgut Changes in Ground Beetles, *Calosomachlorostictum*, under Agricultural Pollution. *Sustainability*. 12: 4805.

- Pez-Osuna, F., Ruiz-Farnndez, A. C., Botello, A. V., Ponce-Velez, G., Osuna-Lopez, J. I., Frias-Espericueta, M. G., Lopez-Lopez, G. and Zazueta-Padilla, H.M. 2002. Concentrations of Selected Trace Metals (Cu, Pb, Zn), Organochlorines (PCBs, HCB) and Total PAHs in Mangrove Oysters from the Pacific Coast of Mexico: An Overview. Mar. Pollut. Bull. 44 : 1296 – 1313.
- Porst, G., Naughton, O., Gill, L., Johnston, P. and Irvine, K. 2012. Adaptation, phenology and disturbance of macroinvertebrates in temporary water bodies. *Hydrobiologia*. 696 : 47–62.
- Poteat, M.D. and Buchwalter, D.B. 2014. Four reasons why traditional metal toxicity testing with aquatic insects is irrelevant. *Environ Sci Technol.* 48 : 887–888.
- Powell, M.I., White, K.M. 1990. Heavy Metal Accumulation by Barnacles and Its Implications for Their Use as Biological Monitors.*Mar. Environ. Res.* 30:91–118.
- Rainbow, R.B. 2007. Trace metal bioaccumulation: Models, metabolic availability and toxicity. *Environ. Entomol.* 33 : 576–582.
- Rawi, S.M., Bakry, F.A. and Al Hazmi, M.A. 2011. Biochemical and histopathological effect of crude extracts on Spodopteralittoralis larvae. *J. Evol. Biol. Res.* 3 : 67–78.
- Sarkar, S.K., Cabral, H., Chatterjee, M., Cardoso, I., Bhattacharya, A.K., Satpathy, K.K. and Alam, M. A. 2008. Biomonitoring of Heavy Metals Using the Bivalve Molluscs in Sunderban Mangrove Wetland, Northeast Coast of Bay of Bengal (India): Possible Risks to Human Health. *Clean*. 36 (2): 187 – 194.
- Shahzad, S., Shahzad, N., Yaqub, A., Sadiq, U., Ahmad, U. And Hussain, M. 2017. Removal of heavy metal lead (pb) (ii) by biosorption process using paper shell mussel (anodontoidesferussacianus). *Wjpmr*. 3 (6).281-288.
- Szefer, P. and Szefer, K., 1990. Metals in molluscs and associated bottom sediments of the southern Baltic. *Helgol. Meeresunters.* 44 : 411–424.
- Thorn, K., Cerrato, R.M. and Rivers, M.L. 1995. Elemental distributions in marine bivalve shells as measured by synchrotron X-Ray-fluorescence. *Biol. Bull.* 188 (1) : 57–67.
- Usero, J., Gonzalez-Regaldo, I. and Gracia. 1997. Trace metals in bivalve molluscs *Ruditapesdecussatus* and *Ruditapesphilippinarum* from the Atlantic coast of Southern Spain. *Environmental International*. 23. 291– 298.
- Vijayaraghavan, K., Winnie, H.Y.N. and Balasubramanian, R. 2011. Biosorption characteristics of crab shell particles for the removal of manganese(II) and zinc(II) from aqueous solutions. *Desalination*. 266 (1-3) : 195-200.
- Walker, K.F. 1992. Ecology of the freshwater mussels in the River Murray. *Australian Water Resources Council Technical Paper*. 63.

- Xie, L., Funk, D.H. and Buchwalter, D.B. 2010. Trophic transfer of Cd from natural periphyton to the grazing mayfly *Centroptilumtriangulifer* in a life cycle test. *Environ Pollut*. 158 : 272–277.
- Yasoshima, M. and Takano, B. 2001. Bradybaenasimilaris (Ferrusac) shell as a biomonitor of copper, cadmium, and zinc. *Bull. Environ. Contam. Toxicol.* 66 (2) : 239–

248.

Zulkifli, S.Z., Ismail, A. and Mohamat-Yusuff, F. 2012. Bioaccumulation of Selected Heavy Metals in Soldier Crabs, Dotillamyctiroides (Decapoda: Ocypodidae) from Bagan Lalang, Selangor, Malaysia. *Acta Biologica Malaysiana*. 94-100.